Abstract—In the work a planar topology for nanoelectronic transducer based on the enzyme glucose oxidase is developed. An experimental study of the possibility of forming nanogaps in planar nanostructures by electromigration is carried out. A technique of implementation of planar nanostructures for nanoelectronic biosensor based on the methods of electron-beam lithography is developed. A technological map of manufacturing of nanogaps between metal electrodes by electromigration of metal atoms on the basis of a computer-controlled process of breaking metal nanowires is developed and realized. The method of the chemical modification of silicon dioxide surface by epoxysilane was developed and the immobilization of the glucose oxidase enzyme on the surface of planar nanostructure for the nanoelectronic transducer through the linker molecules was realized. The registration procedure of the biochemical signals was developed. A change in the functional state of the immobilized ferment of the glucose oxidase enzyme with the presence (oxidation) of glucose in the test solution was demonstrated.

Index Terms—Biosensor, nanotransistor, nanostructures, enzymatic electrochemical sensors, enzymatic activity, environmental monitoring.

I. INTRODUCTION

The biosensors represent the complex analytical devices that use biological materials for "recognition" of certain molecules and give information about their presence and quantity in the form of an electrical signal. The principle of analysis, realized in biosensors, based on the fact that the biological material (enzymes, cells, organelles, immunocomponents) immobilized on physical sensors, produces at the interaction with the analyzed compounds a concentration-dependent electric signal [1].

Generally the operation of the majority of modern biosensors is based on the enzymatic catalysis. A conjugation of the enzymatic catalytic and electrochemical reactions occurring on conductive materials, immersed in an electrolyte solution, allow to develop a lot of biosensors for the determination of various biologically active compounds.

As one of the most common model of biocatalysts for the biosensor analysis the glucose oxidase and alcohol oxidase are used. The glucose oxidase is a highly selective enzyme with in details studied characteristics. The alcohol oxidase from methylotrophic yeast Pichia angusta is capable of catalyzing the oxidation of aliphatic alcohols, and is used in biosensors for analysis [2].

Functionally the biosensors are comparable to sensors of a live organism - the bioreceptors, capable to transform all types of the signals arriving from environment, to the electric ones. Now the most widely biosensors based on enzymes are used. A significant amount of enzymatic biosensors is focused on the analysis of liquids, including biological (for example, for express diagnostics of glucose concentration in blood). Besides, the enzymatic biosensors find application in biotechnology, the food industry, the nature conservation and adjacent areas. They help to solve the various biomedical problems and control the state of the environment (for example, for control of the content of toxic substances) [3].

Any biosensor consists of two basic functional elements: the bioselective element using various biological structures and the physical converter of the signal (transducer) transforming a concentration signal to the electric signal one. The electronic amplification and registration systems are used for reading and recording of the information. As a bioselective element it is possible to use all types of biological structures: enzymes, antibodies, receptors, nucleonic acids and even living cells. In turn the electrochemical converters (electrodes), different optical converters, gravitational, calorimetric, resonant systems can be used as the transducers. All types of bioselective elements can be combined with various transducers. It makes a large variety of various types of biosensors.

As a whole the class of enzymes- oxidases is high-specific in relation to defined substrata. The advantage of this type of a biosensor is first of all in its high selectivity which is defined by specificity of used enzymes and the nature of electrochemical reaction in which the components of enzymatic process participate. On the contrary a systems on the basis of not biological converter, are not selective that is caused by a variety of reasons.

The last achievements in a solution of the problem of miniaturization electronic devices, and also successes in combination of technologies of biology and nanoelectronics allow to develop original versions of nanodevices, in particular, transistors on the basis of nanowires and single molecules that gives an essentially new possibility of use of nanostructures for not only the development of miniature physical devices, but also for the solution of more wide range of tasks in live systems, for example, for production of the miniature biosensor diagnostic units [4].

II. MANUFACTURING TECHNIQUES OF PLANAR NANOSTRUCTURE

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To provide an electrical connection with nanostructures, previously the chip infrastructure of the supplying gold wires conducting in the central region of size 90×90 micrometers was made. The transfer of mask pattern on a substrate was carried out by photolithography. For a photolithography the technology of the double-layer mask (Fig. 1) was used. This technology allows avoiding low-quality border due to contact of sputtering metal with photoresist walls. [5].

The double-layer technology consists of several stages: application of isolating silicon dioxide on a silicon substrate, consecutive application of layers of copolymer and polymer, the UV exposure, processing in toluol and alcohol mixture, a chrome and gold sputtering, mask dissolution in acetone.

![Fig. 1. Comparison single-layer (a) and double-layer (b) of lift-off technology.](image)

A resist PMGI SF6 (MicroChem) was used as the bottom layer of the mask, and Shipley S1805 was used as the top resist layer. The photoresist was exposed by means of installation and exposition system SET MA-750. The double-layer resistive mask was developed in MF-319 developer (Metal Free). As a result, the areas, free from the resist (so-called "windows") appear on a substrate in the exposed places after this intermediate stage. The size of the canopy of the top resist over bottom can be regulated by change of drying temperature of the bottom resist layer.

Into the resulting "windows" in the photoresist a buffer layer of chromium (2 nm) (for adhesion of gold on silicon oxide) and layer of gold (40 nm) were thermally deposited at a pressure of 9×10^{-7} mbar by Leybold L-560. As a result, after removing of photoresist the structure of macroelectrodes (Fig. 2) on the chip was formed.

The measuring nanostructure is disposed in the central area of the chip (90×90 microns) and it was a prototype of system of electrodes for the molecular transistor. The formation of nanostructure was carried out by high resolution lithography with an electron microscope with energy of about 10 keV. In work MicroChem 950 PMMA C2 polymer was used as a top resist layer, and the copolymer MMA(8.5)MAA EL11 was used as the bottom one.

The resulting copolymer film thickness is about 500 nm, and the thickness of the polymer film about 150 nm. The system of measuring electrodes with nanowires in the central part of the chip is shown in Fig. 3.

![Fig. 3. The nanoelectrodes topology of the nanostructure.](image)

The measuring nanoelectrodes for the molecular transistor can be obtained by a breakup of a gold film with forming a gap of nanometer scale [6]. For a breakup of a thin-film nanowire the electromigration technique can be effectively used. This technique requires the ability to set a high density current through a gold nanowire, and control its parameters in real time. By means of electromigration technology the breakup in the center of a nanowire was made and the technological nanogaps for fixing of enzyme molecules were formed (Fig. 4).

![Fig. 4. The nanogap image. Size of image is 2×2 μm².](image)

III. THE TECHNOLOGY OF AN ENZYME OF MOLECULES IMMOBILIZATION

In the experiments three enzymatic preparations of glucose oxidase (GOD) were used: GOD synthesized of Penicillium admetzii LF F-2044.1 (activity of 41.4 unit/mg), GOD of Penicillium funiculosum 46.1 (activity of 36.4 unit/mg), and GOD of Aspergillus niger (activity of 265 unit/mg) (Valeant Pharmaceuticals, the USA). At preliminary research of properties of glucose oxidases there was checked the influence of active acidity in the range of pH 2-12 (0.01 M the universal buffer) and temperatures in the range of 20-80°C. [7].

For an immobilization of glucose oxidase enzyme on the composite substrate, which was a silicon covered by silicon dioxide, the cross-link method of a protein groups through a epoxysilane groups to a hydroxyl groups on a the surface of a silicon dioxide was applied.

![Fig. 2. The topology of supplying electrodes on the chip. Size of chip is 10x10 mm².](image)
The research of the model of a nanoelectronic transducer on the basis of planar nanostructure with built-in molecules of enzymes was preceded by preliminary experiments with use of converters of electrochemical type. There were used the substrates with gold nanoelectrodes. There was investigated the connection of electrodes to glucose oxidase enzymes which deposited in the form of solution, and also immobilized with use of glutaraldehyde (an enzyme put in microquantities). The obtained results were compared further to data on electronic transport processes of the transfer of a charge in nanostructures.

The electronic properties of a nanoelectronic transducer were investigated in buffer solution both in absence and at addition of a glucose oxidase. It is shown that reaction to glucose is absent on a control sample (Fig. 8), and an electronic response of the biosensor to 10 mM solution of glucose is present on the nanoelectronic structure modified by enzyme (Fig. 9).
V. CONCLUSION

Thus, in work the production technology of the model of the nanoelectronic transducer on the basis of planar nanostructure with built-in enzymatic complexes is designed. The technology of an immobilization of molecules of enzyme on nanostructure electrodes is elaborated. The technique of biochemical signals recording is developed. The change of a functional state of the immobilized enzyme of glucose oxidase in the presence of glucose (oxidation) in the test solution is demonstrated.

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